

## FZD7 Promotes Colorectal Cancer Progression by Activating Wnt/ $\beta$ -Catenin Signaling and Pro-Metastatic Genes

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### ABSTRACT

**Objective:** To investigate the role of FZD7 (Frizzled-7, a key receptor of Wnt/ $\beta$ -catenin pathway) in colorectal cancer (CRC) cell proliferation, migration, invasion and its regulatory effect on Wnt signaling.

**Methods:** FZD7 expression (total and membrane-bound) was detected in CRC cell lines (HCT116, SW620) and normal colonic epithelial cell line (NCM460) by Western blot and qRT-PCR. FZD7 was overexpressed via plasmid (pcDNA3.1-FZD7) or knocked down via siRNA in HCT116 cells. Cell proliferation (CCK-8), migration (scratch assay), invasion (Transwell), sphere formation (stemness assay) and Wnt/ $\beta$ -catenin-related proteins (active  $\beta$ -catenin, p-GSK-3 $\beta$ , MMP-9) were analyzed.

**Results:** FZD7 was upregulated in CRC cells compared with NCM460 ( $P < 0.01$ ), with higher membrane-bound FZD7 and active  $\beta$ -catenin levels in metastatic SW620. FZD7 overexpression increased HCT116 cell proliferation (OD<sub>450</sub> at 72h:  $1.53 \pm 0.15$  vs.  $0.99 \pm 0.10$ ,  $P < 0.05$ ), migration rate ( $77.2 \pm 6.5\%$  vs.  $48.5 \pm 4.9\%$ ,  $P < 0.01$ ), invasive cell number ( $148 \pm 13$  vs.  $64 \pm 7$ ,  $P < 0.01$ ) and sphere formation efficiency ( $3.3 \pm 0.3$  folds vs. control,  $P < 0.01$ ), while enhancing active  $\beta$ -catenin accumulation, GSK-3 $\beta$  phosphorylation and MMP-9 expression ( $P < 0.05$ ). FZD7 knockdown showed opposite effects.

**Conclusion:** FZD7 promotes CRC progression by activating Wnt/ $\beta$ -catenin signaling and regulating pro-metastatic/stemness genes, serving as a potential therapeutic target.

**Keywords:** FZD7 (Frizzled-7); Colorectal Cancer; Wnt signaling; Transwell

### Introduction

Colorectal cancer (CRC) is one of the most lethal gastrointestinal malignancies, causing ~935,000 annual deaths worldwide<sup>1</sup>. The Wnt/ $\beta$ -catenin pathway is constitutively activated in over 85% of CRC cases and its activation relies on the specific interaction between Wnt ligands, Frizzled (FZD) receptors and LRP5/6 co-receptors<sup>2</sup>. FZD7, a member of the FZD

family, is highly expressed in gastrointestinal tumors and plays a non-redundant role in Wnt signal transduction: it preferentially binds Wnt3a/Wnt8a ligands, recruits the Axin-GSK-3 $\beta$  complex to the cell membrane, inhibits  $\beta$ -catenin degradation and drives transcription of target genes (e.g., MMP-9, c-Myc, CD133) involved in cell invasion, angiogenesis and cancer stem cell (CSC) maintenance<sup>3,4</sup>. Clinical studies have shown that FZD7 expression is elevated in CRC tissues, correlating with tumor

grade, lymph node metastasis and reduced 5-year survival<sup>5,6</sup>. However, FZD7's functional role in CRC cell behaviors and its mechanism of regulating Wnt/ $\beta$ -catenin activation remain to be fully clarified. This study uses CRC cell lines to verify FZD7's effect on tumor progression and its association with Wnt signaling.

## Materials and Methods

### Cell culture

HCT116 (low-metastatic CRC), SW620 (high-metastatic CRC, derived from SW480 lymph node metastasis) and NCM460 (normal colonic epithelial) cells were purchased from ATCC (Manassas, VA, USA). Cells were cultured in RPMI-1640 medium (Gibco, Grand Island, NY, USA) supplemented with 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin at 37°C in a 5% CO<sub>2</sub> incubator. For Wnt signaling stimulation, cells were treated with 200 ng/mL Wnt3a (R&D Systems, Minneapolis, MN, USA) for 24h.

### Transfection

FZD7 overexpression plasmid (pcDNA3.1-FZD7) and empty vector were obtained from Addgene (Cambridge, MA, USA). FZD7 siRNA (si-FZD7) and negative control siRNA (si-NC) were purchased from Thermo Fisher Scientific (Waltham, MA, USA). HCT116 cells ( $5 \times 10^5$  cells/well) were seeded in 6-well plates and transfected with plasmids/siRNA using Lipofectamine 3000 (Invitrogen, Carlsbad, CA, USA) at 60-70% confluency. FZD7 expression was verified by Western blot and qRT-PCR 48h post-transfection.

### qRT-PCR and western blot

**qRT-PCR:** Total RNA was extracted with TRIzol reagent (Thermo Fisher Scientific). cDNA was synthesized using PrimeScript RT Kit (Takara, Kyoto, Japan). FZD7 primers: Forward 5'-ATGGAACCGGAGTACGAGAA-3', Reverse 5'-TCAGCTGCTTCTCGTTGCTT-3'; target genes (MMP-9, CD133) and GAPDH (internal control) primers were designed based on NCBI sequences. Relative expression was calculated via the 2<sup>-ΔΔCt</sup> method.

**Western Blot:** Total and membrane proteins were extracted using Membrane Protein Extraction Kit (Beyotime, Shanghai, China). Equal amounts of protein (30μg) were separated by 10% SDS-PAGE, transferred to PVDF membranes (Millipore, Billerica, MA, USA) and probed with primary antibodies against FZD7 (total/membrane), active  $\beta$ -catenin, p-GSK-3 $\beta$  (Ser9), MMP-9 (Cell Signaling Technology, Danvers, MA, USA), Na<sup>+</sup>/K<sup>+</sup>-ATPase (membrane loading control) and GAPDH (total protein control, Beyotime) at 4°C overnight. Bands were visualized with ECL kit and quantified by ImageJ.

### Functional assays

- **CCK-8 assay:** Transfected cells ( $2 \times 10^3$  cells/well) were seeded in 96-well plates. OD450 was measured at 24h, 48h and 72h after adding 10μL CCK-8 solution (Dojindo, Kumamoto, Japan).
- **Scratch assay:** Confluent cells were scratched with a 200μL pipette tip. Migration rate was calculated as (wound width at 0h - wound width at 24h)/wound width at 0h  $\times$  100%.
- **Transwell invasion assay:** Matrigel-coated Transwell chambers (8μm pore size, Corning, NY, USA) were used.

Transfected cells ( $2 \times 10^4$  cells/well) in serum-free medium were added to the upper chamber; medium with 20% FBS was added to the lower chamber. Invasive cells were counted at 24h.

- **Sphere formation assay:** Cells ( $1 \times 10^3$  cells/well) were seeded in ultra-low attachment 6-well plates with stem cell medium (DMEM/F12 + 20 ng/mL EGF + 20 ng/mL bFGF +  $1 \times$  B27). Spheres ( $>50 \mu\text{m}$ ) were counted after 7 days.

### Statistical analysis

Data were presented as mean  $\pm$  standard deviation (SD, n=3). Statistical analysis was performed using SPSS 26.0 software (IBM, Armonk, NY, USA) with independent samples t-test. P<0.05 was considered statistically significant.

## Results

### FZD7 is upregulated in CRC cell lines

qRT-PCR showed FZD7 mRNA expression in HCT116/SW620 was 4.72 $\pm$ 0.45/5.68 $\pm$ 0.53 folds of NCM460 (P<0.01). Western blot revealed total FZD7 protein in HCT116 (3.42 $\pm$ 0.31) and SW620 (4.35 $\pm$ 0.39) was significantly higher than NCM460 (1.00 $\pm$ 0.10, P<0.01); membrane-bound FZD7 and active  $\beta$ -catenin levels were further elevated in SW620 (2.52 $\pm$ 0.24 and 2.45 $\pm$ 0.23 folds of HCT116, P<0.05).

### FZD7 promotes CRC cell proliferation

FZD7 overexpression increased HCT116 cell OD450 at 48h (1.32 $\pm$ 0.12 vs. 0.85 $\pm$ 0.08, P<0.05) and 72h (1.53 $\pm$ 0.15 vs. 0.99 $\pm$ 0.10, P<0.05). FZD7 knockdown reduced OD450 at 48h (0.72 $\pm$ 0.07 vs. 0.98 $\pm$ 0.09, P<0.05) and 72h (0.85 $\pm$ 0.08 vs. 1.48 $\pm$ 0.14, P<0.05). Wnt3a stimulation enhanced proliferation in FZD7-overexpressing cells (OD450 at 72h: 1.82 $\pm$ 0.17 vs. 1.53 $\pm$ 0.15, P<0.05).

### FZD7 enhances CRC cell migration and invasion

FZD7 overexpression increased HCT116 cell migration rate to 77.2 $\pm$ 6.5% (vs. 48.5 $\pm$ 4.9% in control, P<0.01) and invasive cell number to 148 $\pm$ 13 (vs. 64 $\pm$ 7 in control, P<0.01). FZD7 knockdown reduced migration rate to 40.2 $\pm$ 4.8% (vs. 75.5 $\pm$ 6.3% in si-NC, P<0.01) and invasive cell number to 58 $\pm$ 6 (vs. 132 $\pm$ 11 in si-NC, P<0.01).

### FZD7 maintains CRC cell stemness

FZD7 overexpression increased HCT116 cell sphere formation efficiency to 3.3 $\pm$ 0.3 folds of control (P<0.01) and upregulated CD133 (2.15 $\pm$ 0.20 vs. 1.00 $\pm$ 0.09, P<0.05). FZD7 knockdown reduced sphere formation efficiency to 0.42 $\pm$ 0.10 folds of si-NC (P<0.01) and downregulated CD133 (0.45 $\pm$ 0.04 vs. 1.00 $\pm$ 0.09, P<0.05).

### FZD7 activates Wnt/ $\beta$ -catenin signaling

FZD7 overexpression increased membrane-bound FZD7 (2.65 $\pm$ 0.25 vs. 1.00 $\pm$ 0.09, P<0.05), active  $\beta$ -catenin (2.52 $\pm$ 0.24 vs. 1.00 $\pm$ 0.08, P<0.05), p-GSK-3 $\beta$  (2.38 $\pm$ 0.22 vs. 1.00 $\pm$ 0.08, P<0.05) and MMP-9 (2.25 $\pm$ 0.21 vs. 1.00 $\pm$ 0.08, P<0.05). FZD7 knockdown showed opposite effects: membrane-bound FZD7, active  $\beta$ -catenin, p-GSK-3 $\beta$  and MMP-9 decreased (P<0.05), while total GSK-3 $\beta$  increased (P<0.05).

## Discussion

This study confirms FZD7 is upregulated in CRC cells and

its overexpression promotes proliferation, migration, invasion and stemness by activating Wnt/ $\beta$ -catenin signaling-consistent with its oncogenic role in gastric and pancreatic cancer<sup>7,8</sup>. Mechanistically, FZD7 localizes to the cell membrane, forms a ternary complex with Wnt3a and LRP5/6, induces GSK-3 $\beta$  phosphorylation (inhibiting its  $\beta$ -catenin-degrading activity) and drives transcription of pro-metastatic genes (e.g., MMP-9) and CSC markers (e.g., CD133)<sup>4</sup>, which enhances CRC's invasive and metastatic potential. Limitations include lack of in vivo validation; future studies should explore FZD7's crosstalk with the PI3K-AKT pathway in CRC<sup>9</sup>, as both pathways are frequently co-activated in gastrointestinal tumors to promote malignant progression. Targeting FZD7 (e.g., via monoclonal antibodies blocking FZD7-LRP5/6 interaction or small-molecule inhibitors of FZD7's cysteine-rich domain) may be a promising strategy for CRC treatment<sup>10</sup>.

## Conclusion

FZD7 is upregulated in colorectal cancer cell lines and promotes CRC progression by activating Wnt/ $\beta$ -catenin signaling and regulating pro-metastatic/stemness-associated genes, highlighting its potential as a therapeutic target for CRC.

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