

## CSNK1A1 Inhibits Colorectal Cancer Progression by Suppressing Canonical Wnt/ $\beta$ -Catenin Signaling via $\beta$ -Catenin Phosphorylation

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### ABSTRACT

**Objective:** To investigate the role of CSNK1A1 (casein kinase 1 alpha 1, a key regulator of canonical Wnt/ $\beta$ -catenin pathway) in colorectal cancer (CRC) cell proliferation, migration, invasion and its regulatory effect on Wnt signaling.

**Methods:** CSNK1A1 expression was detected in CRC cell lines (HCT116, SW480) and normal colonic epithelial cell line (NCM460) by Western blot and qRT-PCR. CSNK1A1 was overexpressed via plasmid (pcDNA3.1-CSNK1A1) or knocked down via siRNA in HCT116 cells. Cell proliferation (CCK-8), migration (scratch assay), invasion (Transwell), sphere formation (stemness assay) and canonical Wnt-related proteins ( $\beta$ -catenin, p- $\beta$ -catenin Ser45, AXIN1, c-Myc) were analyzed.

**Results:** CSNK1A1 was downregulated in CRC cells compared with NCM460 ( $P < 0.01$ ), with lower expression in metastatic SW480. CSNK1A1 overexpression decreased HCT116 cell proliferation ( $OD_{450}$  at 72h:  $0.72 \pm 0.07$  vs.  $1.02 \pm 0.10$ ,  $P < 0.05$ ), migration rate ( $40.2 \pm 4.7\%$  vs.  $49.5 \pm 5.0\%$ ,  $P < 0.01$ ), invasive cell number ( $55 \pm 6$  vs.  $68 \pm 7$ ,  $P < 0.01$ ) and sphere formation efficiency ( $0.40 \pm 0.04$  folds vs. control,  $P < 0.01$ ), while enhancing  $\beta$ -catenin Ser45 phosphorylation (promoting degradation), increasing AXIN1 stability and downregulating c-Myc ( $P < 0.05$ ). CSNK1A1 knockdown showed opposite effects.

**Conclusion:** CSNK1A1 functions as a tumor suppressor in CRC by inhibiting canonical Wnt/ $\beta$ -catenin signaling via  $\beta$ -catenin phosphorylation, serving as a potential therapeutic target for restoring pathway homeostasis.

**Keywords:** CSNK1A1 (casein kinase 1 alpha 1); Transwell; Wnt signaling

### Introduction

Colorectal cancer (CRC) is a leading cause of cancer-related mortality globally, with ~935,000 annual deaths<sup>1</sup>. The canonical Wnt/ $\beta$ -catenin pathway is constitutively activated in over 85% of CRC cases and its activity is tightly regulated by sequential phosphorylation of  $\beta$ -catenin—first by CSNK1A1 (Ser45), then by GSK-3 $\beta$  (Thr41/Ser37/Ser33)—to trigger ubiquitination

and degradation<sup>2,3</sup>. CSNK1A1, a member of the casein kinase 1 family, is a critical upstream kinase in this process: it not only initiates  $\beta$ -catenin phosphorylation but also stabilizes the AXIN1-containing “destruction complex” by phosphorylating AXIN1, further enhancing  $\beta$ -catenin degradation<sup>4,5</sup>. Clinical studies have shown that CSNK1A1 is frequently downregulated or mutated in CRC tissues, correlating with tumor stage, lymph

node metastasis and reduced 5-year survival<sup>6,7</sup>. However, CSNK1A1's functional role in CRC cell behaviors (especially pathway suppression) and its mechanism of regulating Wnt/ $\beta$ -catenin homeostasis remain to be fully clarified. This study uses CRC cell lines to verify CSNK1A1's tumor-suppressive effect and its association with canonical Wnt signaling.

## Materials and Methods

### Cell culture

HCT116 (low-metastatic CRC), SW480 (high-metastatic CRC) and NCM460 (normal colonic epithelial) cells were purchased from ATCC (Manassas, VA, USA). Cells were cultured in RPMI-1640 medium (Gibco, Grand Island, NY, USA) supplemented with 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin at 37°C in a 5% CO<sub>2</sub> incubator. For Wnt pathway activation, cells were treated with 200 ng/mL Wnt3a protein (R&D Systems, Minneapolis, MN, USA) for 24h.

### Transfection

CSNK1A1 overexpression plasmid (pcDNA3.1-CSNK1A1) and empty vector were obtained from Addgene (Cambridge, MA, USA). CSNK1A1 siRNA (si-CSNK1A1) and negative control siRNA (si-NC) were purchased from Thermo Fisher Scientific (Waltham, MA, USA). HCT116 cells ( $5 \times 10^5$  cells/well) were seeded in 6-well plates and transfected with plasmids/siRNA using Lipofectamine 3000 (Invitrogen, Carlsbad, CA, USA) at 60-70% confluency. CSNK1A1 expression was verified by Western blot and qRT-PCR 48h post-transfection.

### qRT-PCR and western blot

**qRT-PCR:** Total RNA was extracted with TRIzol reagent (Thermo Fisher Scientific). cDNA was synthesized using PrimeScript RT Kit (Takara, Kyoto, Japan). CSNK1A1 primers: Forward 5'-ATGGAACCGGAGTACGAGAA-3', Reverse 5'-TCAGCTGCTTCTCGTTGCTT-3'; target genes (c-Myc, Cyclin D1) and GAPDH (internal control) primers were designed based on NCBI sequences. Relative expression was calculated via the  $2^{-\Delta\Delta Ct}$  method.

**Western blot:** Total and nuclear proteins were extracted using Nuclear Extraction Kit (Beyotime, Shanghai, China). Equal amounts of protein (30 $\mu$ g) were separated by 10% SDS-PAGE, transferred to PVDF membranes (Millipore, Billerica, MA, USA) and probed with primary antibodies against CSNK1A1,  $\beta$ -catenin (total/nuclear), p- $\beta$ -catenin (Ser45), AXIN1, c-Myc (Cell Signaling Technology, Danvers, MA, USA), Lamin B1 (nuclear loading control) and GAPDH (total protein control, Beyotime) at 4°C overnight. Bands were visualized with ECL kit and quantified by ImageJ.

### Functional assays

- **CKK-8 Assay:** Transfected cells ( $2 \times 10^3$  cells/well) were seeded in 96-well plates. OD450 was measured at 24h, 48h and 72h after adding 10 $\mu$ L CKK-8 solution (Dojindo, Kumamoto, Japan).
- **Scratch Assay:** Confluent cells were scratched with a 200 $\mu$ L pipette tip. Migration rate was calculated as (wound width at 0h - wound width at 24h)/wound width at 0h  $\times$  100%.
- **Transwell Invasion Assay:** Matrigel-coated Transwell chambers (8 $\mu$ m pore size, Corning, NY, USA) were used. Transfected cells ( $2 \times 10^4$  cells/well) in serum-free medium

were added to the upper chamber; medium with 20% FBS was added to the lower chamber. Invasive cells were counted at 24h.

- **Sphere Formation Assay:** Cells ( $1 \times 10^3$  cells/well) were seeded in ultra-low attachment 6-well plates with stem cell medium (DMEM/F12 + 20 ng/mL EGF + 20 ng/mL bFGF +  $1 \times$  B27). Spheres ( $>50 \mu$ m) were counted after 7 days.

### Statistical analysis

Data were presented as mean  $\pm$  standard deviation (SD, n=3). Statistical analysis was performed using SPSS 26.0 software (IBM, Armonk, NY, USA) with independent samples t-test. P<0.05 was considered statistically significant.

## Results

### CSNK1A1 is downregulated in CRC cell lines

qRT-PCR showed CSNK1A1 mRNA expression in HCT116/SW480 was  $0.45 \pm 0.04/0.32 \pm 0.03$  folds of NCM460 (P<0.01). Western blot revealed CSNK1A1 protein in HCT116 ( $0.42 \pm 0.04$ ) and SW480 ( $0.28 \pm 0.03$ ) was significantly lower than NCM460 ( $1.00 \pm 0.10$ , P<0.01); nuclear  $\beta$ -catenin levels were inversely elevated in SW480 ( $2.95 \pm 0.27$  folds of HCT116, P<0.05), while p- $\beta$ -catenin (Ser45) was reduced ( $0.35 \pm 0.04$  folds of HCT116, P<0.05).

### CSNK1A1 inhibits CRC cell proliferation

CSNK1A1 overexpression decreased HCT116 cell OD450 at 48h ( $0.85 \pm 0.08$  vs.  $1.05 \pm 0.09$ , P<0.05) and 72h ( $0.72 \pm 0.07$  vs.  $1.02 \pm 0.10$ , P<0.05). CSNK1A1 knockdown increased OD450 at 48h ( $1.28 \pm 0.12$  vs.  $1.05 \pm 0.09$ , P<0.05) and 72h ( $1.52 \pm 0.14$  vs.  $1.02 \pm 0.10$ , P<0.05). Wnt3a stimulation partially reversed CSNK1A1-induced proliferation inhibition (P<0.05).

### CSNK1A1 Reduces CRC cell migration and invasion

CSNK1A1 overexpression decreased HCT116 cell migration rate to  $40.2 \pm 4.7\%$  (vs.  $49.5 \pm 5.0\%$  in control, P<0.01) and invasive cell number to  $55 \pm 6$  (vs.  $68 \pm 7$  in control, P<0.01). CSNK1A1 knockdown increased migration rate to  $65.8 \pm 6.2\%$  (vs.  $49.5 \pm 5.0\%$  in si-NC, P<0.01) and invasive cell number to  $92 \pm 8$  (vs.  $68 \pm 7$  in si-NC, P<0.01).

### CSNK1A1 suppresses CRC cell stemness

CSNK1A1 overexpression decreased HCT116 cell sphere formation efficiency to  $0.40 \pm 0.04$  folds of control (P<0.01) and downregulated CD44 ( $0.45 \pm 0.04$  vs.  $1.00 \pm 0.09$ , P<0.05). CSNK1A1 knockdown increased sphere formation efficiency to  $2.3 \pm 0.2$  folds of si-NC (P<0.01) and upregulated CD44 ( $2.15 \pm 0.20$  vs.  $1.00 \pm 0.09$ , P<0.05).

### CSNK1A1 inactivates canonical Wnt/ $\beta$ -catenin signaling

CSNK1A1 overexpression increased p- $\beta$ -catenin (Ser45) ( $2.65 \pm 0.25$  vs.  $1.00 \pm 0.09$ , P<0.05) and AXIN1 stability ( $1.85 \pm 0.17$  vs.  $1.00 \pm 0.08$ , P<0.05), while reducing nuclear  $\beta$ -catenin ( $0.48 \pm 0.04$  vs.  $1.00 \pm 0.09$ , P<0.05) and c-Myc ( $0.52 \pm 0.05$  vs.  $1.00 \pm 0.08$ , P<0.05). CSNK1A1 knockdown showed opposite effects: p- $\beta$ -catenin (Ser45) and AXIN1 decreased (P<0.05), while nuclear  $\beta$ -catenin and c-Myc increased (P<0.05), indicating inhibited  $\beta$ -catenin degradation.

## Discussion

This study confirms CSNK1A1 is downregulated in CRC

cells and its overexpression exerts tumor-suppressive effects by inhibiting proliferation, migration, invasion and stemness-consistent with its role in gastric and pancreatic cancer<sup>8,9</sup>. Mechanistically, CSNK1A1 initiates  $\beta$ -catenin phosphorylation at Ser45, a prerequisite for subsequent GSK-3 $\beta$ -mediated phosphorylation and degradation; it also stabilizes AXIN1 to reinforce the destruction complex, thereby suppressing nuclear translocation of  $\beta$ -catenin and transcription of pro-oncogenic genes (e.g., c-Myc)<sup>5</sup>. Limitations include lack of in vivo validation; future studies should explore CSNK1A1's interaction with Wnt co-receptors (e.g., LRP6) in CRC<sup>10</sup>, as CSNK1A1 also phosphorylates LRP6 to modulate Wnt pathway activation. Restoring CSNK1A1 activity (e.g., via small-molecule activators or kinase agonists) may be a promising strategy for CRC treatment.

## Conclusion

CSNK1A1 is downregulated in colorectal cancer cell lines and inhibits CRC progression by suppressing canonical Wnt/ $\beta$ -catenin signaling via  $\beta$ -catenin phosphorylation and AXIN1 stabilization, highlighting its potential as a therapeutic target for restoring pathway homeostasis in CRC.

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